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It can be done

# Assessment of Ionic and Neutral PFAS Using Newly Developed Equilibrium Regimen Passive Sampler



Analytical  
Services



Health  
Assessment



Novel  
Chemistries



Remediation  
& Treatment



Site  
Characterization



Toxicology

# The need

- PFAS are hydrophobic and oleophobic and display surfactant properties.
- Uses: fire-fighting foams (AFFF), non-staining carpets, water-resistant paper, and more.
- Growing evidence for adverse PFAS health effects.
- 2016 EPA guidance of 70 ng/L; States are mandating stricter regulations/guidelines of PFAS and PFOS.
- 100s of contaminated sites need characterization and remediation.
- Environmental concentration can fluctuate depending on precipitation.
- Cross-contamination makes sampling a challenge.



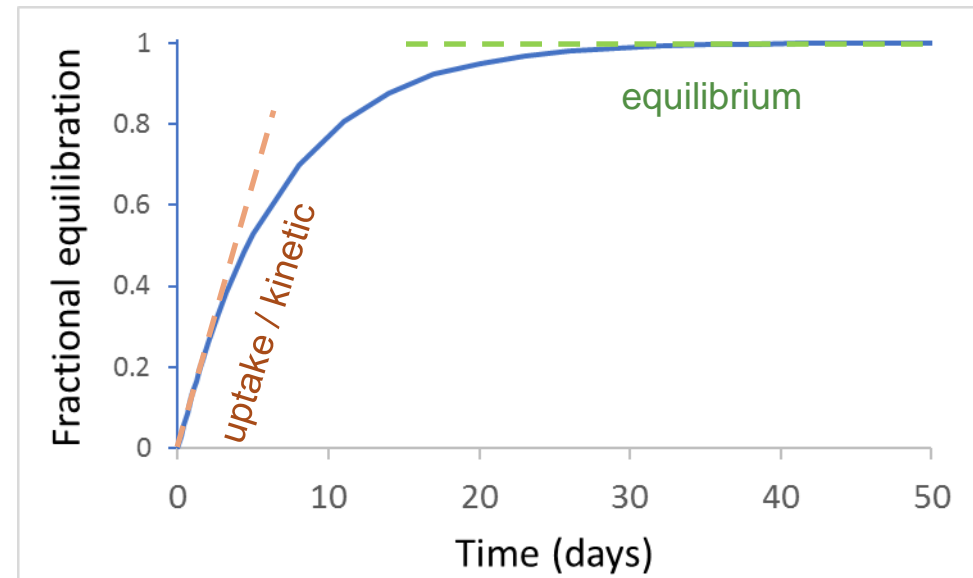
# Why passive sampling?

## General

- Passive samplers for hydrophobic organic contaminants
  - Improved detection limits
  - Time-integrated results
  - Insights into bioavailability
  - Smaller sample size
  - Less cross-contamination risk

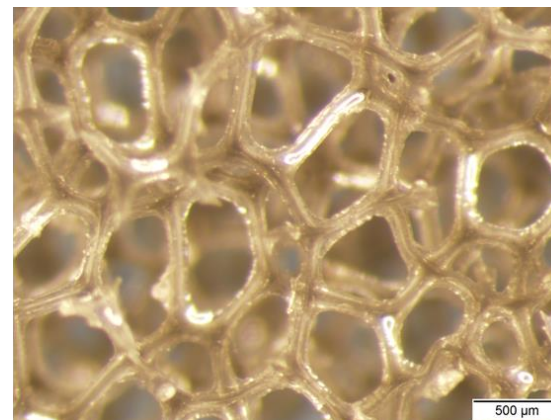
## PFAS considerations

- Wide range of properties
- The sorbent is key!
- Kinetic vs. equilibrium regimen



# PFAS Insight™

- Equilibrium regimen sampler
- After initial testing, down-selected to 2 types of sorbents for further investigation
  - Both sorbents are open-cell polymers
  - 45 pores per inch; SA ~0.4–0.5 m<sup>2</sup>/g
- Tested for 15 analytes
  - non-ionic (precursors) and anionic (sulfonates and carboxylate species)
- Analytical method
  - AB Sciex QTRAP 5500 Triple Quadrupole MS
  - Method compliant with DoD QSM 5.1 Table B-15 requirements



Analyte	# of C
<b>Precursors</b>	
6:2 FTS	6
8:2 FTS	8
NMeFOSAA	8
NEtFOSAA	8
<b>Sulfonates</b>	
PFBS	4
PFH <sub>x</sub> S	6
PFOS	8
PFDS	10
<b>Carboxylates</b>	
PFH <sub>x</sub> A	6
PFHpA	7
PFOA	8
PFNA	9
PFDA	10
PFUnA	11
PFDoA	12

# PFAS uptake kinetics



1 x 1 cm piece of sorbents

0.01 M NaCl background electrolyte

Spike PFAS target analytes  
Initial conc. 10 - 100  $\mu\text{g L}^{-1}$



Triplicates for all treatments plus duplicated blanks and controls



Analysis on LC-MS/MS

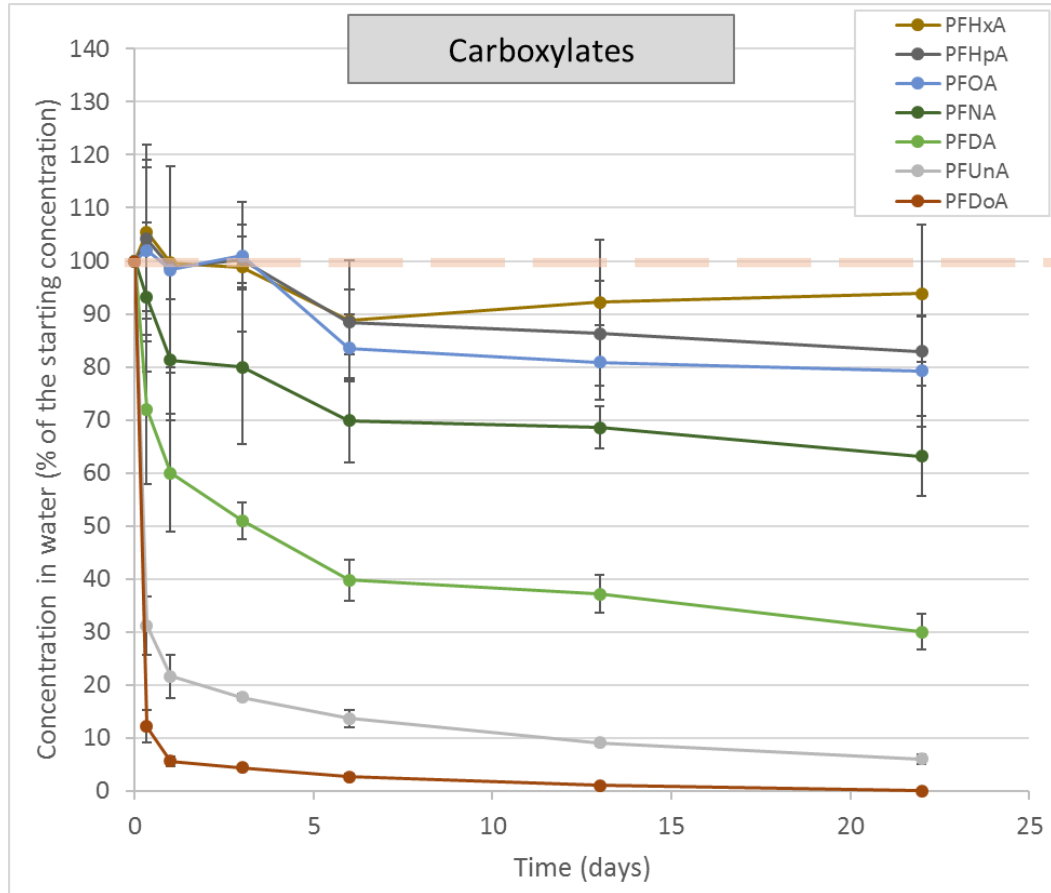
Sample dilution  
Surrogates & Internal standard spiked



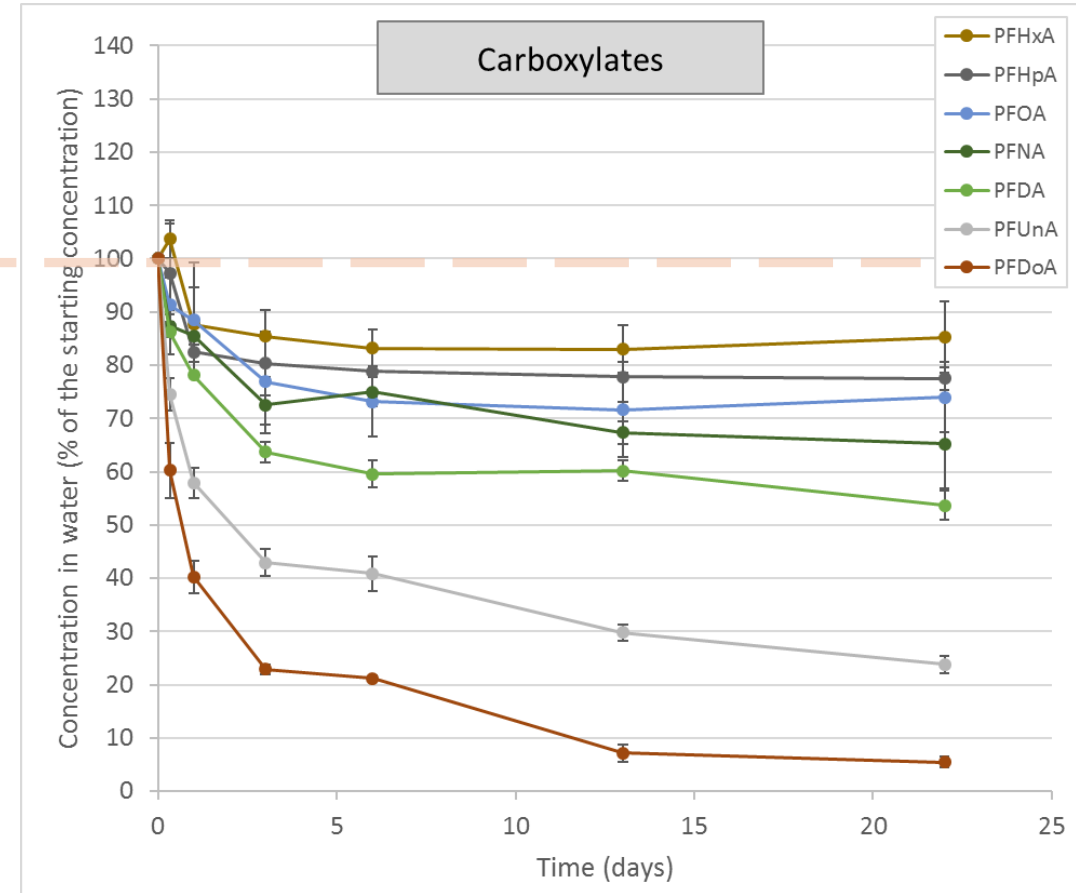
Shaken at 130 rpm,  $23 \pm 1^\circ\text{C}$  and sampled over 0-22 d

# PFAS uptake – water concentration vs. time

Sorbent 1



Sorbent 2



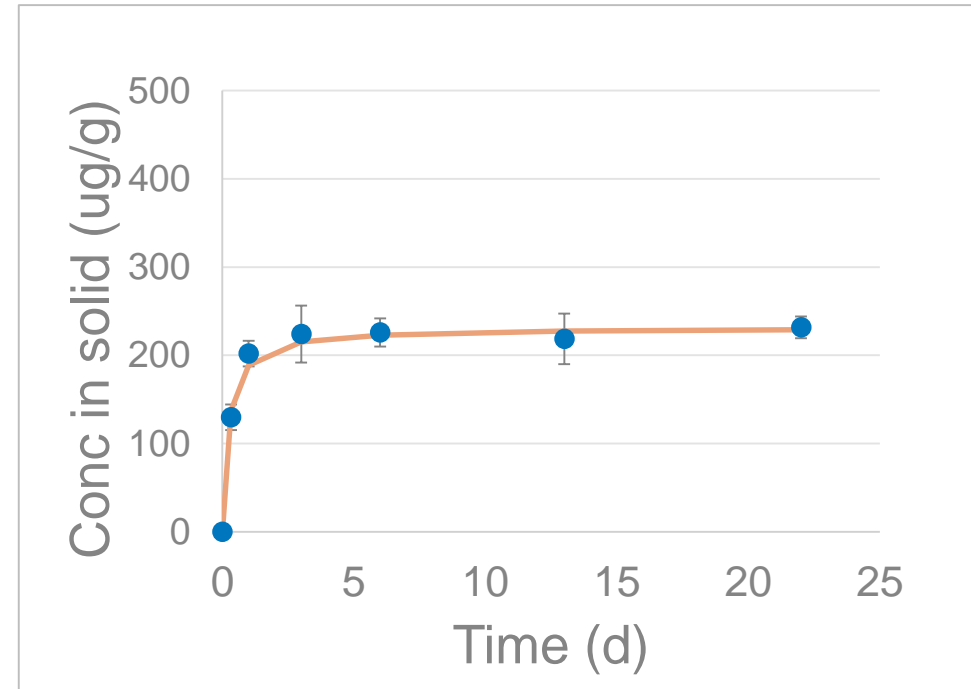
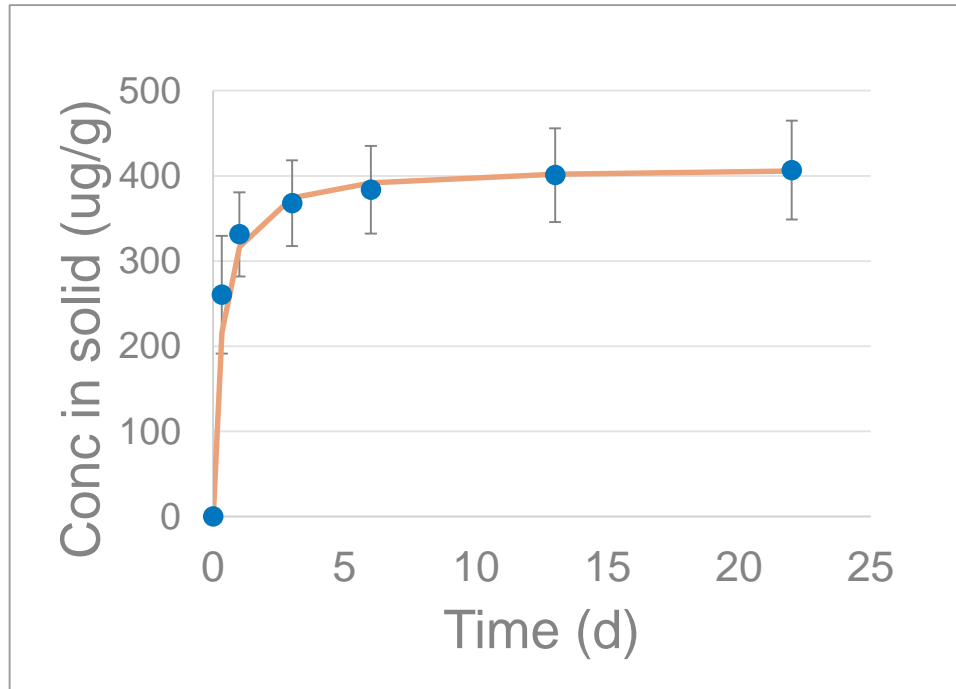
# PFAS uptake – pseudo-second order kinetics

Non-linear:  $C_s = \frac{C_e^2 k t}{1 + C_e t}$

Linear:  $\frac{t}{C_s} = \frac{1}{kC_e^2} + \frac{t}{C_e} = \frac{1}{v_0} + \frac{t}{C_e}$

Sorbent 1

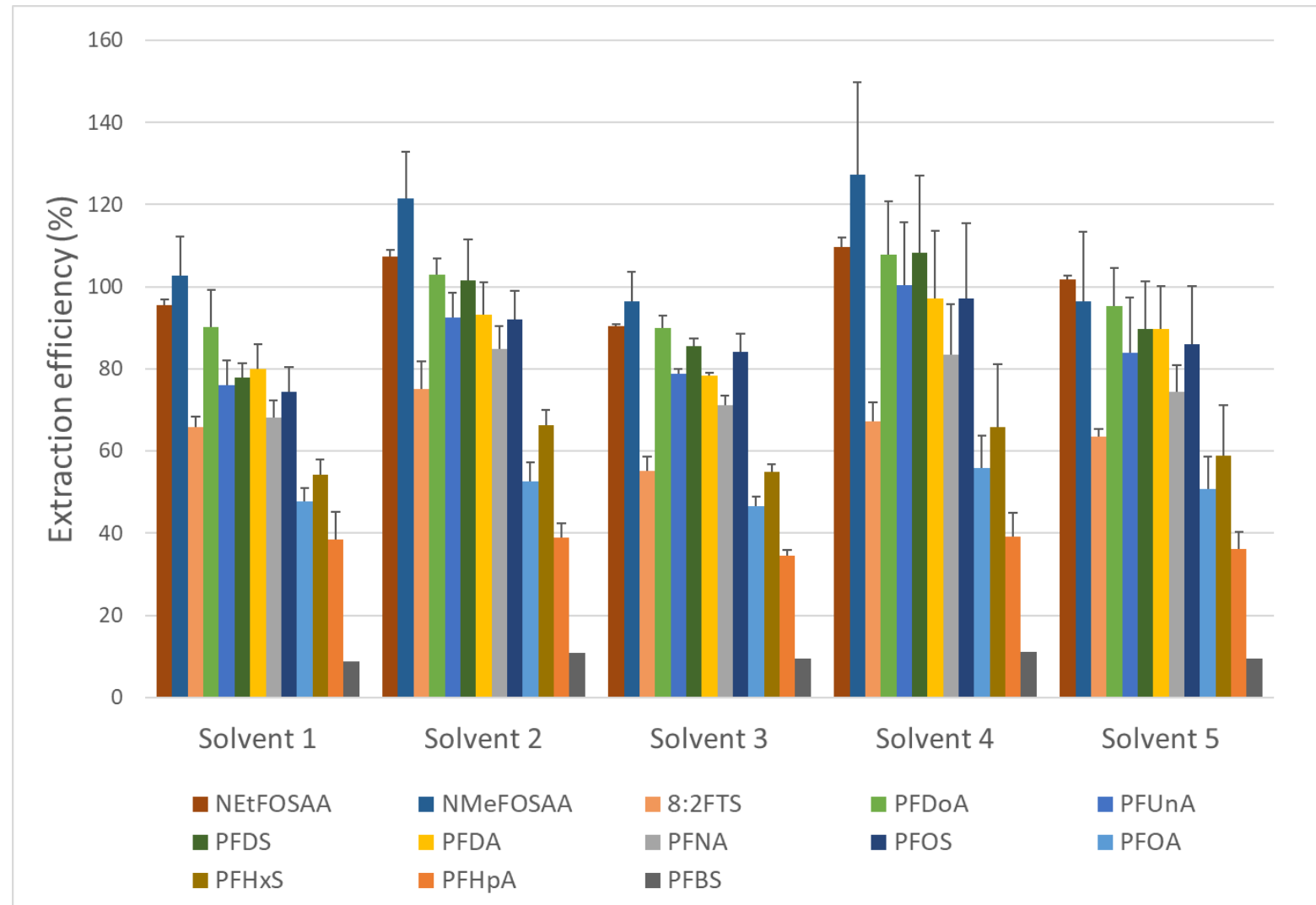
Sorbent 2



PFOS

# Extraction of spent sorbent

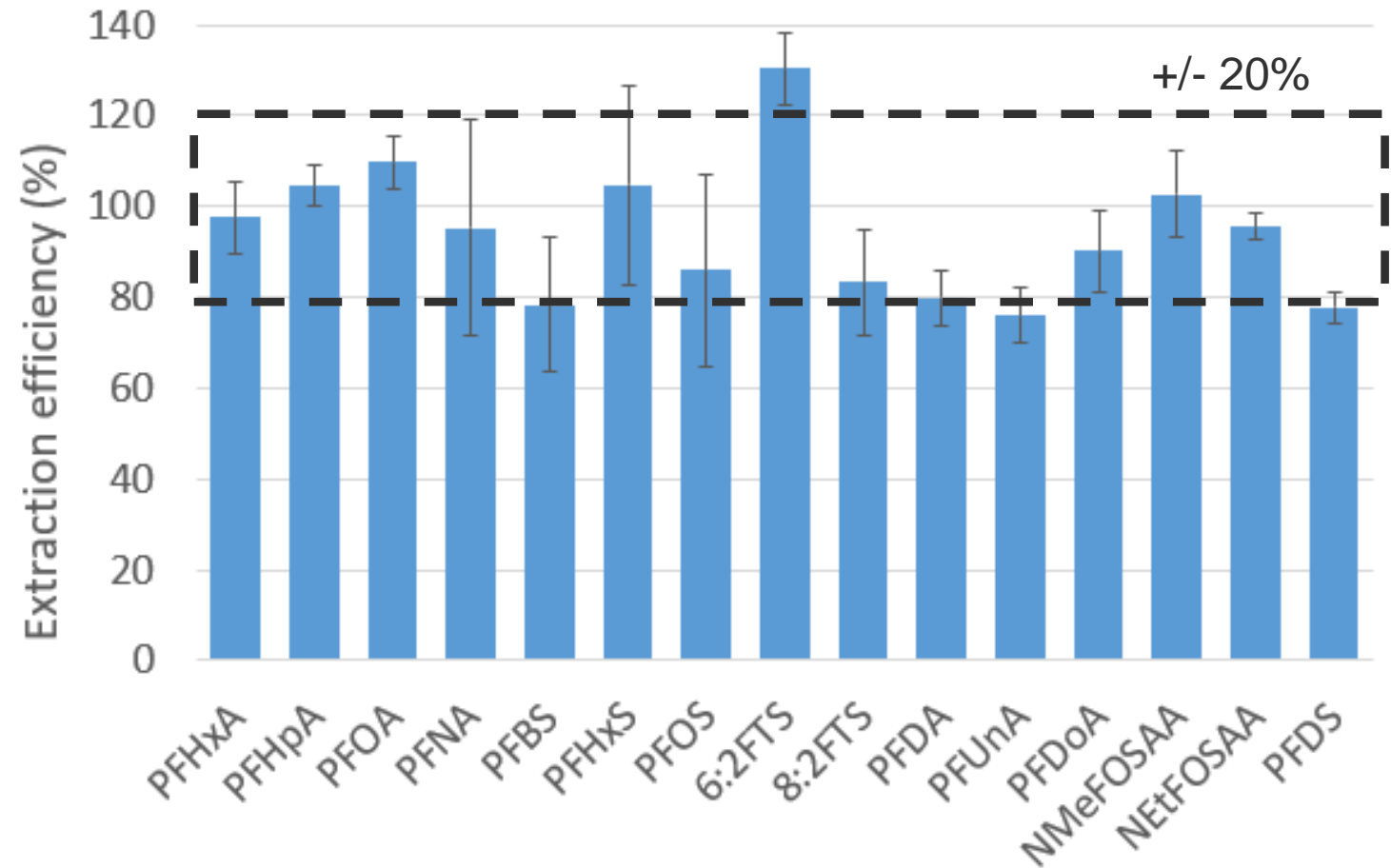
- Experimental design
  - Five pH-modified organic solvents.
  - Three consecutive extractions with varying extraction time and combination of agitation and or/ sonication.
  - Extracts were combined and sampled; no preconcentration.



# Extraction of spent sorbent

## • Conclusions

- Some solvents degraded Sorbent 2.
- Sonication did not cause significant improvement of analyte recovery.
- Overnight + two short extractions achieve great recoveries, with the minimum of 76% of the initial amount (average of 94%).



# Adsorption isotherm experiments

- 1 x 1 cm of sorbent (~0.025 g) added to 125 mL of PFAS solution with background electrolyte (0.01M NaCl).
- Analytes concentrations range from 0.05 to 500 µg/L except PFDS, NMeFoSAA and NEtFOSAA which were present at ~0.1 x the concentration of other analytes.
- Agitated at room temperature for 21 days.
- Duplicates or triplicates plus positive and negative controls.
- Amount adsorbed determined from water sampling at initiation and termination.
- Results modeled using Langmuir and Freundlich equations.

$$q = q_{max} \frac{K_L C}{1 + K_L C}$$

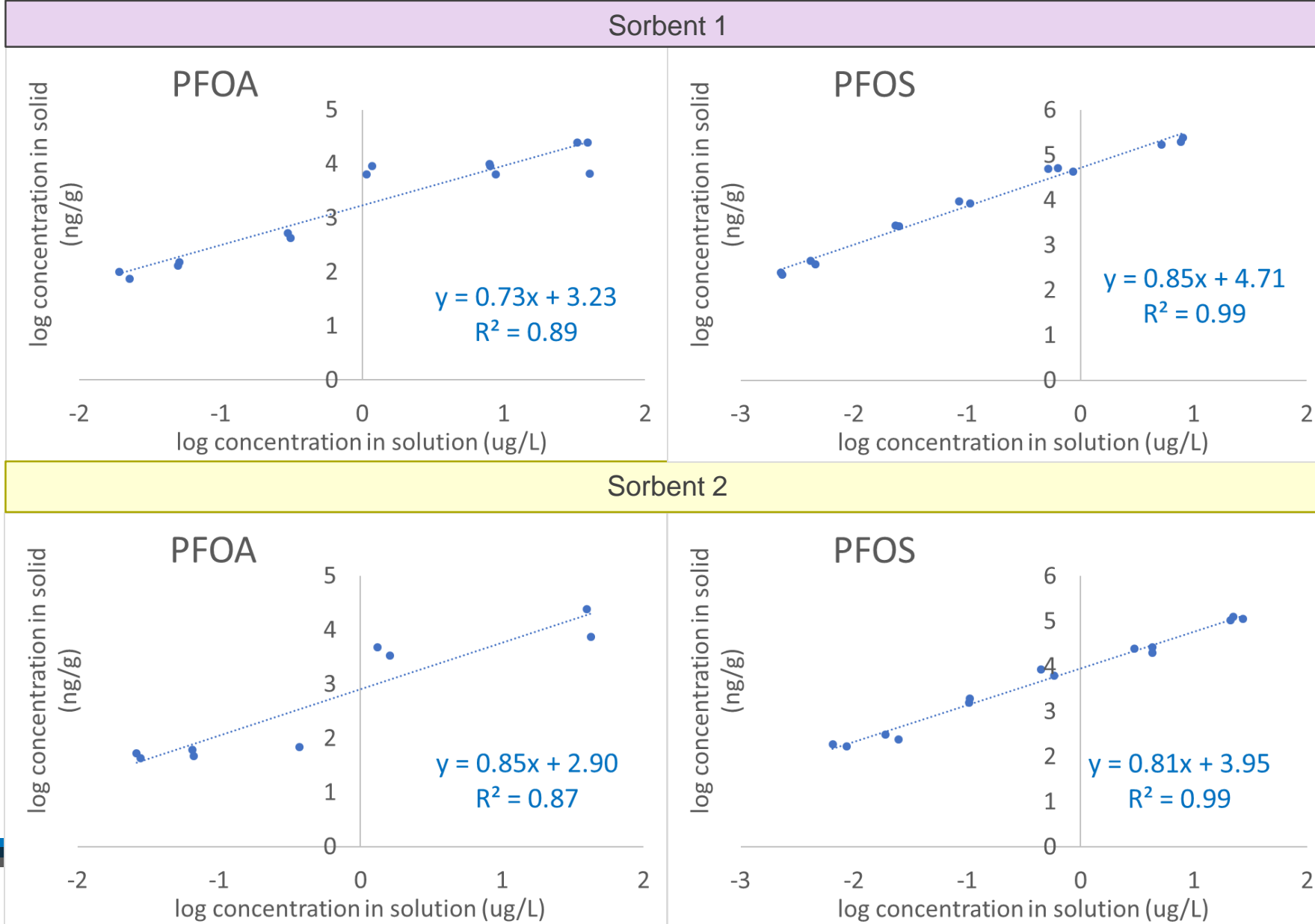
←

$$q = K_F C^n$$

←



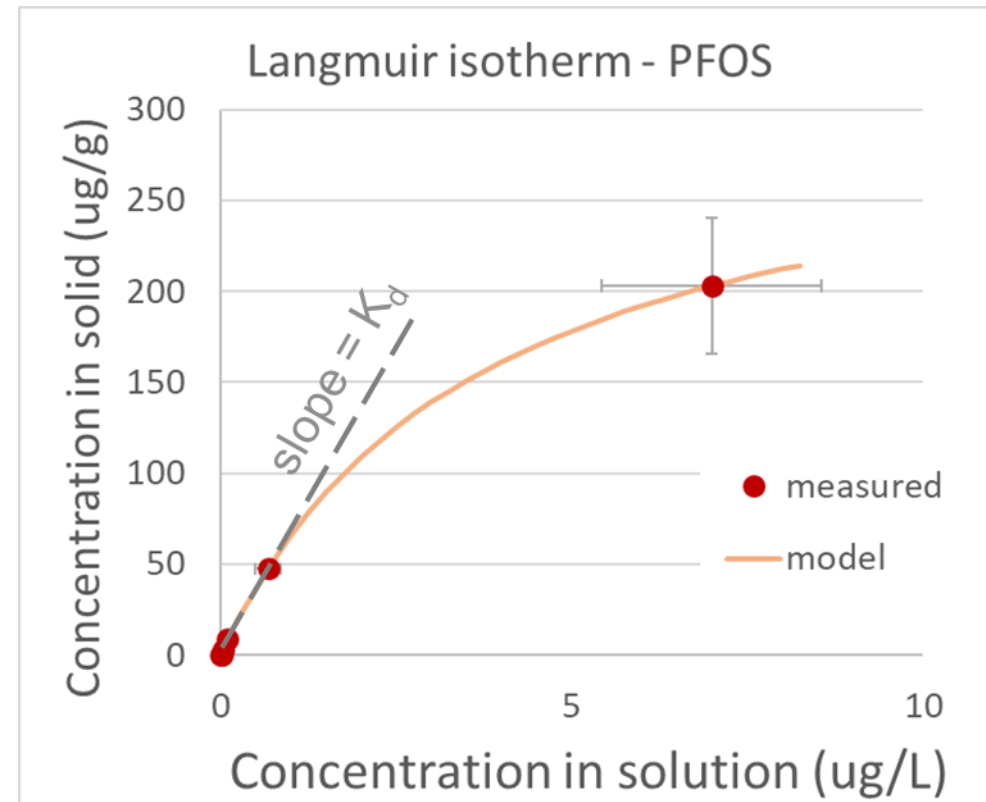
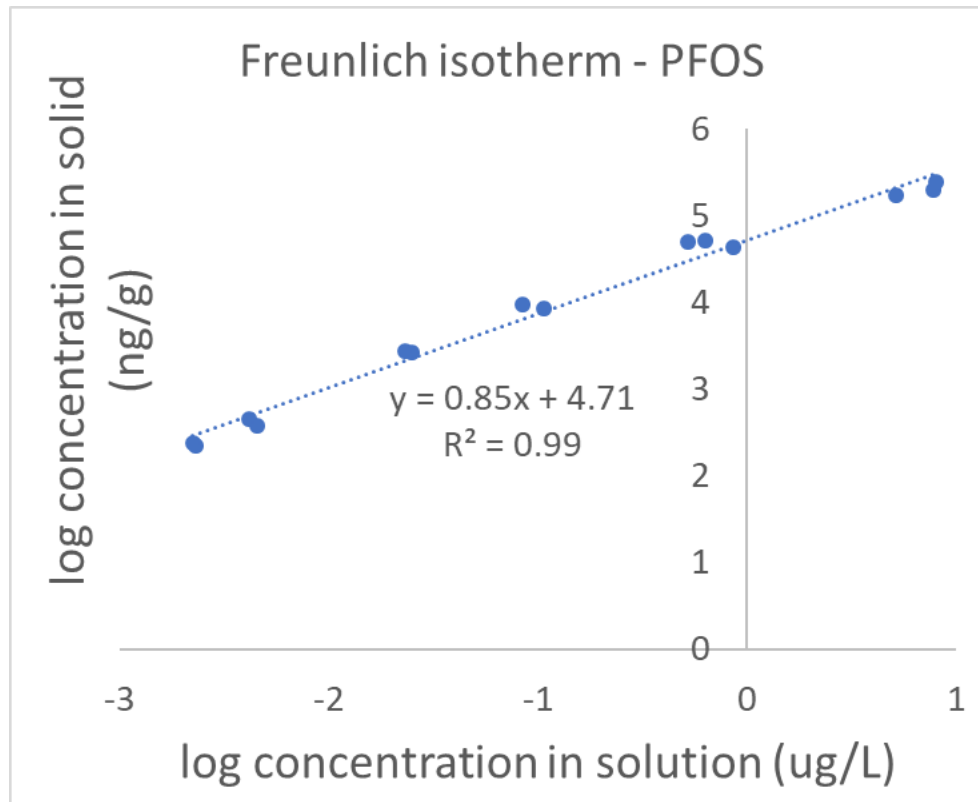
# Adsorption isotherms



# Determination of the solid-solution partition coefficient ( $K_d$ )

$$K_d = \frac{C_{solid}}{C_{solution}}$$

Typical units: L/kg

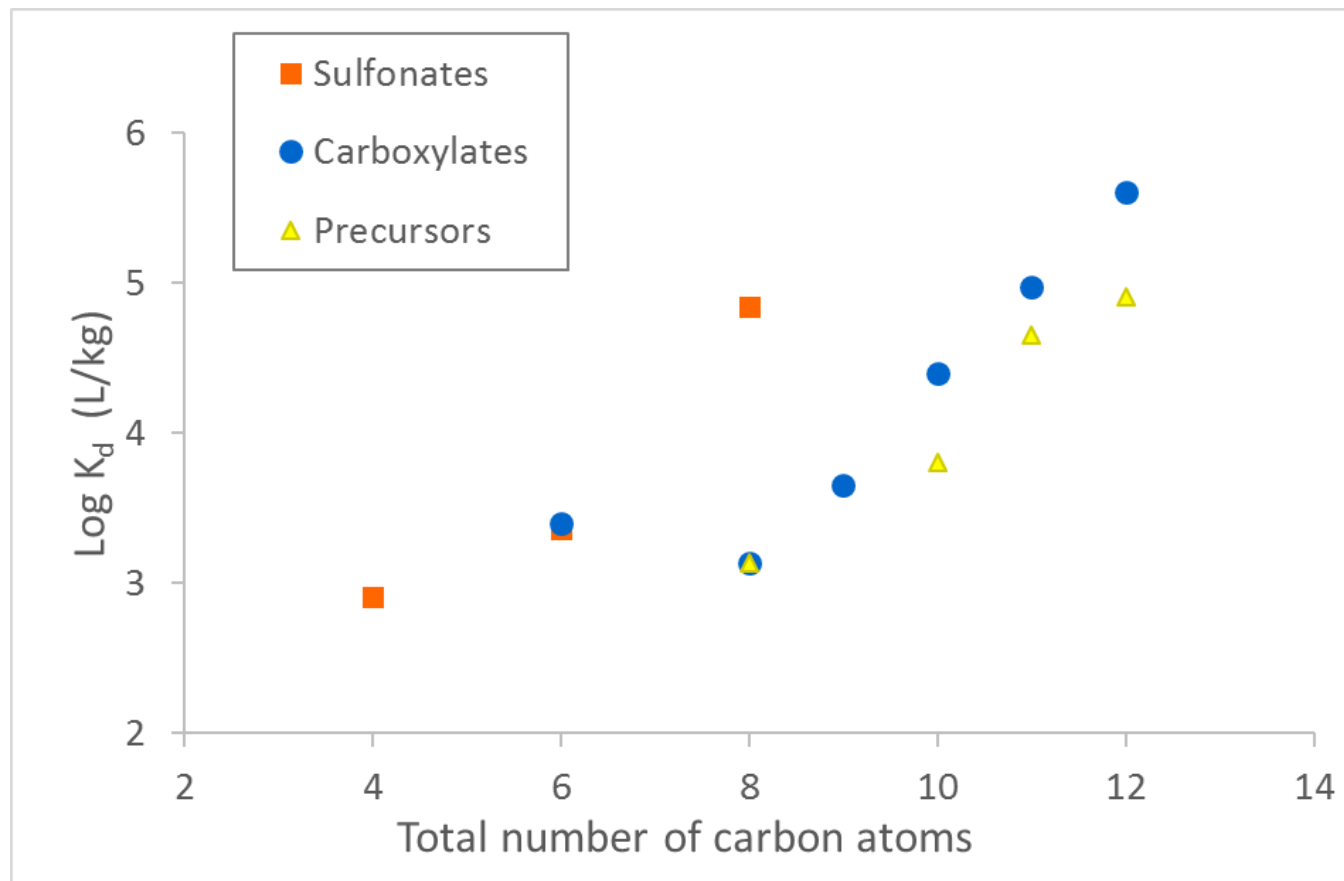


# Partition coefficients ( $K_d$ s)

$$C_w = \frac{C_s}{K_d}$$

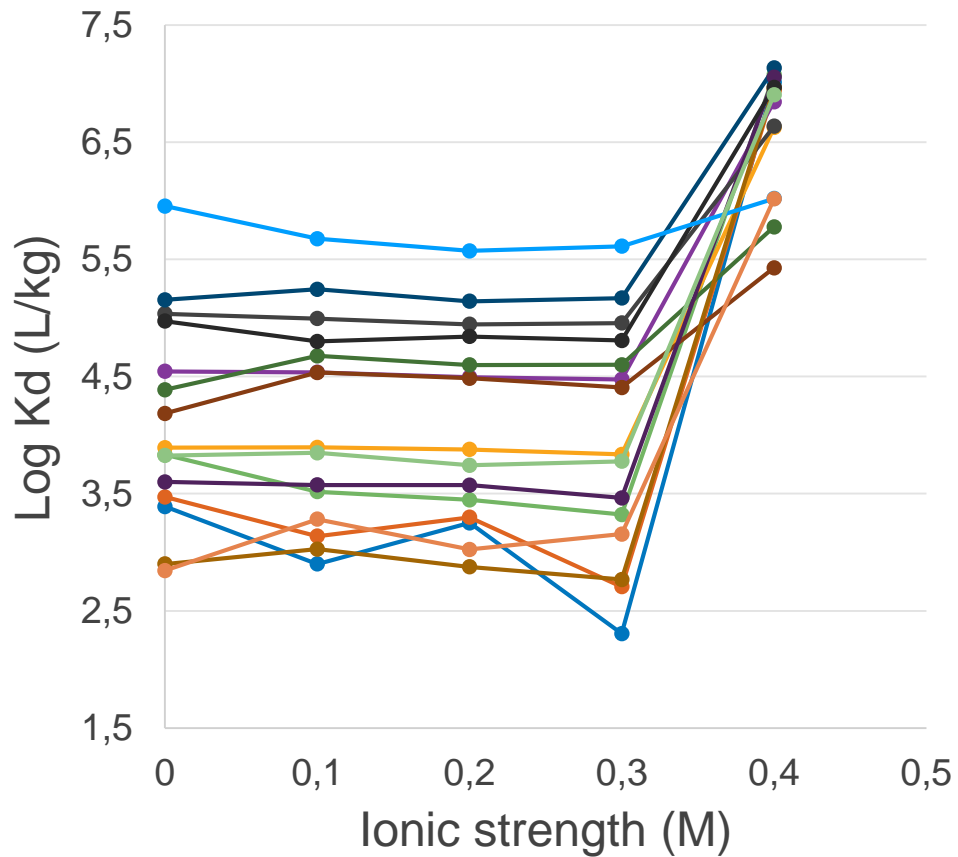
Analyte	Total # of carbons	$K_d$ Sorbent 1	Log $K_d$ Sorbent 1
<b>Precursors</b>			
6:2 FTS	8	1,380	3.14
8:2 FTS	10	6,415	3.81
NMeFOSAA	11	44,716	4.65
NEtFOSAA	12	80,145	4.90
<b>Sulfonates</b>			
PFBS	4	813	2.91
PFHxS	6	2,277	3.36
PFOS	8	69,709	4.84
PFDS	10	NA	NA
<b>Carboxylates</b>			
PFHxA	6	2,529	3.40
PFHpA	7	NA	NA
PFOA	8	1,355	3.13
PFNA	9	4,501	3.65
PFDA	10	25,253	4.40
PFUnA	11	93,725	4.97
PFDoA	12	400,567	5.60

Sorbent 1

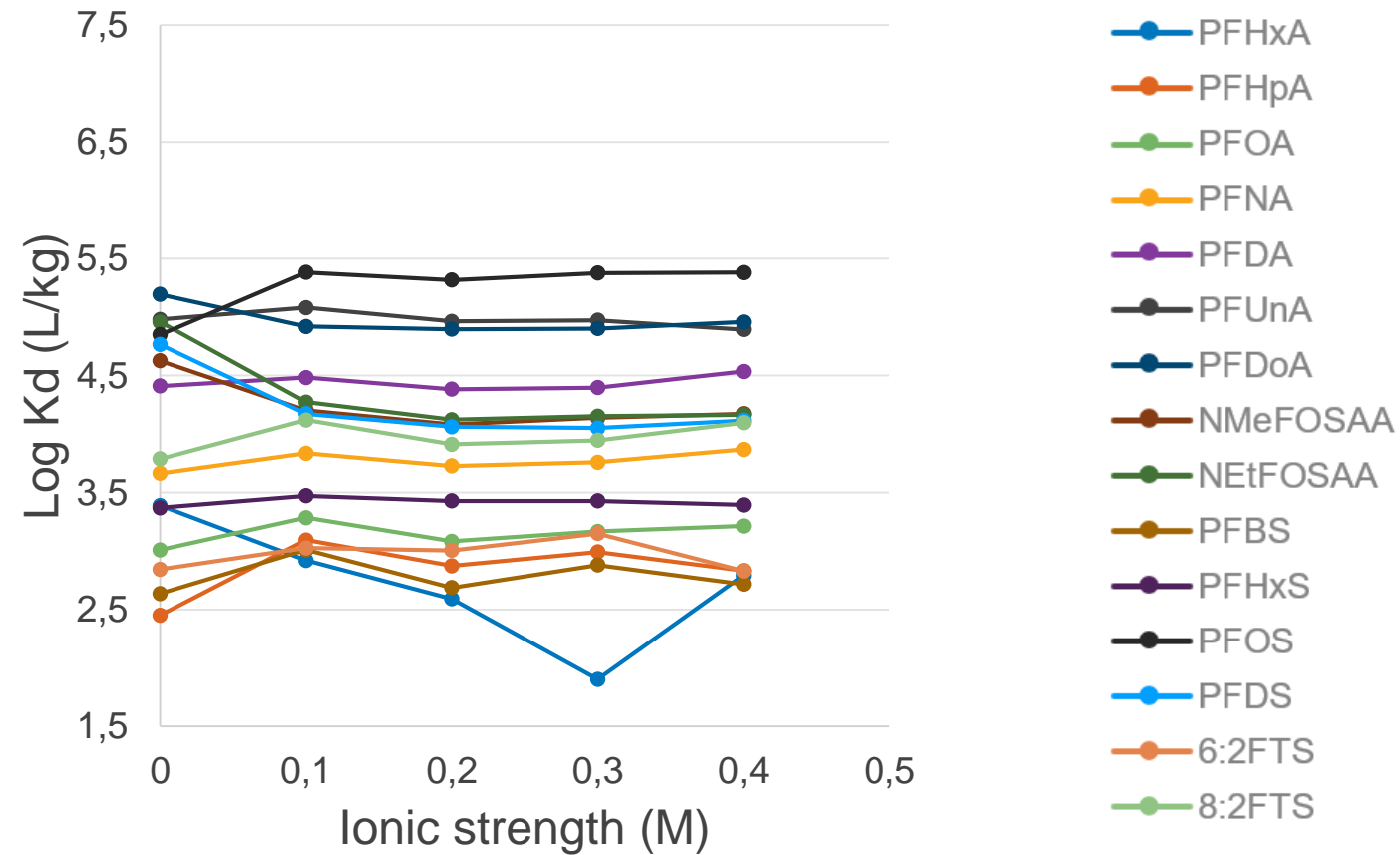


# Effect of Ionic Strength on $K_d$

Starting PFAS concentration = 2 ug/L



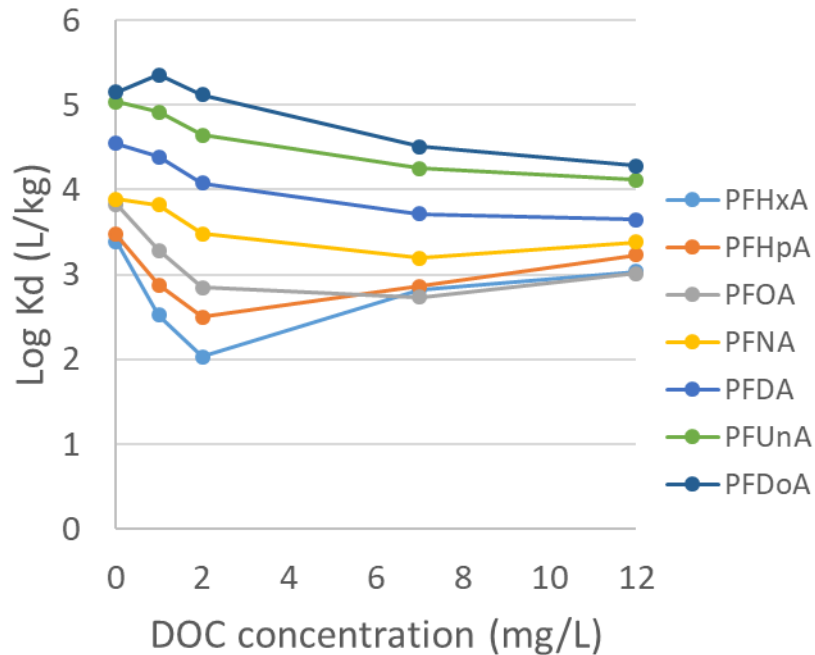
Starting PFAS concentration = 10 ug/L



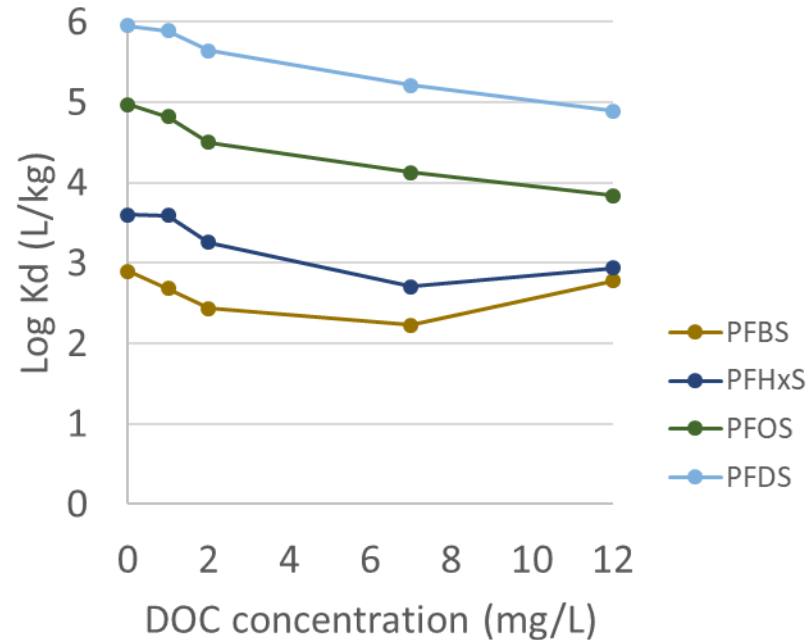
# Effect of Dissolved Organic Carbon (DOC) on $K_d$

Starting PFAS concentration = 2 ug/L

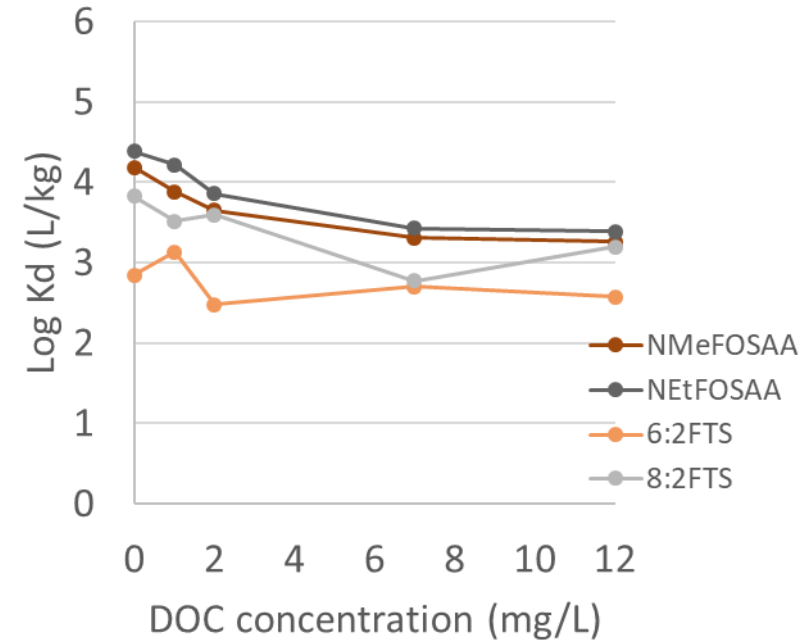
### Carboxylates



### Sulfonates

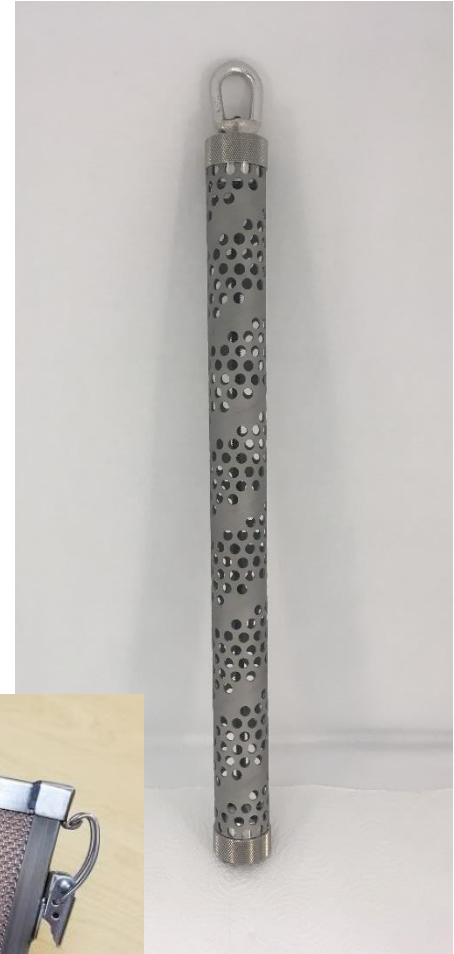


### Precursors



# Hardware Design and Field Deployments

- Two types of hardware were designed to meet the needs of both surface water and groundwater deployments.
- First round of field demonstration-validation PFAS Insight™ samplers retrieved from the field last week and submitted for analysis.
- More field samples to be collected in the near future to allow performance assessment.



# Summary

## Adsorption

- Equilibrium achieved in < 1 week for most analytes.
- Strong preconcentration of anionic and neutral PFAS.
- Adsorption strongest and fastest for longer chains, sulfonates, and precursors.

## Extraction efficiency

- Extraction efficiencies were similar for all solvents tested.
- Extraction procedures optimized to achieve an average of 94% recovery with one overnight and two short extractions.

## Matrix influence

- Increasing ionic strength did not affect partitioning of PFAS except for low PFAS concentration and highest ionic strength (0.4 M) treatment, where increase in  $K_d$  was observed.
- The effect of DOC correlated with PFAS chain length, but generally caused decrease in  $K_d$  by up to ~ 1 log unit.

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For more information on PFAS at Battelle visit  
[www.battelle.org/PFAS](http://www.battelle.org/PFAS)